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Assessing Drivers of Coastal Primary Production in Northern Marguerite Bay, Antarctica

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The coastal ocean of the climatically-sensitive west Antarctic Peninsula is experiencing changes in the physical and (photo)chemical properties that strongly affect the phytoplankton. Consequently, a shift from diatoms, pivotal in the Antarctic food web, to more mobile and smaller flagellates has been observed. We seek to identify the main drivers behind primary production (PP) without any assumptions beforehand to obtain the best possible model of PP. We employed a combination of field measurements and modeling to discern and quantify the influences of variability in physical, (photo)chemical, and biological parameters on PP in northern Marguerite Bay. Field data of high-temporal resolution (November 2013–March 2014) collected at a long-term monitoring site here were combined with estimates of PP derived from photosynthesis-irradiance incubations and modeled using mechanistic and statistical models. Daily PP varied greatly and averaged $1,764 \text{ mg C m}^{-2} \text{ d}^{-1}$ with a maximum of $6,908 \text{ mg C m}^{-2} \text{ d}^{-1}$ after the melting of sea ice and the likely release of diatoms concentrated therein. A non-assumptive random forest model (RF) with all possibly relevant parameters (M_{RFmax}) showed that variability in PP was best explained by light availability and chlorophyll *a* followed by physical (temperature, mixed layer depth, and salinity) and chemical (phosphate, total nitrogen, and silicate) water column properties. The predictive power from the relative abundances of diatoms, cryptophytes, and haptophytes (as determined by pigment fingerprinting) to PP was minimal. However, the variability in PP due to changes in species composition was most likely underestimated due to the contrasting strategies of these phytoplankton groups as we observed significant negative relations between PP and the relative abundance of flagellates groups. Our reduced model (M_{RFmin}) showed how light availability, chlorophyll *a*, and total nitrogen concentrations can be used to obtain the best estimate of PP ($R^2 = 0.93$). The resulting estimates from our models suggest summer PP to have been between 214.4 and 176.1 g C m^{-2} . Through the employment of a modeling technique without any assumptions apart from a representative sampling strategy, we showed and estimated how PP in this climatically sensitive and changing region can best be predicted and described.

Keywords: Antarctica, sea ice, primary production, carbon, phytoplankton, diatoms, haptophytes, cryptophytes

INTRODUCTION

The Western Antarctic Peninsula (WAP) is a region where changes in climate have been most pronounced over recent decades, with strong variability superposed on a long-term warming trend (Vaughan et al., 2003; Meredith and King, 2005; Turner et al., 2005, 2016). These changes include shifting patterns in wind direction and speed, a strong reduction sea ice extent and marked glacial retreat (Marshall et al., 2006; Montes-Hugo et al., 2008; Stammerjohn et al., 2008; Rignot et al., 2013; Turner et al., 2013; Stammerjohn and Maksym, 2017). As a result, the physical characteristics of adjacent marine waters are changing. Increasing wind speeds and the diminution of sea ice have resulted in deepening of mixed layers along the northern WAP region (Montes-Hugo et al., 2009). Moreover, the decrease in sea ice extent removes an important source of freshwater which would otherwise enhance salinity stratification of the water column upon melting in spring. In contrast, an increase in glacial melting is observed, due to enhanced upwelling of relatively warm Circumpolar Deep Water (Cook et al., 2016). These changes may have significant consequences for irradiance and nutrient availability. On the one hand, the increase in glacial meltwater promotes stratification and consequently improves light availability. On the other, more frequent and/or deeper mixing could increase the availability of macro- and micro-nutrients, whereas glacial meltwater is generally poor in macronutrients but potentially rich in trace metals such as iron (Gerringa et al., 2012). This interplay between climate change related impacts is likely to have an effect on both the composition and productivity of the phytoplankton community.

The classical Antarctic food web revolves around highly productive sea ice edge blooms consisting of diatoms, where krill can feed and transfer energy up the food chain (reviewed in Ducklow et al., 2012a). These blooms are instigated by the melting of sea ice and increase in light availability after the polar winter. With the appearance of deeper mixed layers, the phytoplankton community has reoriented itself toward higher proportions of smaller flagellates such as haptophytes (generally *Phaeocystis*) and cryptophytes (Arrigo et al., 1999; Montes-Hugo et al., 2009; Kozłowski et al., 2011; Trimborn et al., 2015; Rozema et al., 2017a). In the WAP region, this shift in phytoplankton community seems further fueled by the replacement of sea ice melt by glacial melt as the major source of freshwater (Moline et al., 2004; Mendes et al., 2013). While diatoms are still the major contributor to phytoplankton biomass, cryptophytes, and haptophytes are increasing in the coastal WAP region (Saba et al., 2014; Rozema et al., 2017a). Our current understanding suggests an association of haptophytes with more unstable water columns, while cryptophytes show increased abundances when these water columns are restabilizing due to glacial meltwater injection.

The occurring shift in the phytoplankton community seems to coincide with changes in the primary productivity (PP) of the coastal WAP ecosystem. Physical properties of the water column, for example stability, influence PP directly and indirectly. A less stable water column means less available light for photosynthesis, but also promotes growth of different phytoplankton groups. Phytoplankton biomass accumulation is

limited in years following winters with a more unstable water column due to low sea ice cover during winter (Venables et al., 2013). This potential effect is further strengthened due to the changing species composition as productivity per biomass unit of small flagellates is different to that of diatoms (reviewed in Petrou et al., 2016). For example, *Phaeocystis antarctica* is thought to be more productive per chlorophyll a (Chl-a) and per captured photon than *Fragilariopsis cylindrus*, a small diatom associated with the sea ice edge (Arrigo et al., 2010; Alderkamp et al., 2012a). In contrast, data suggest a decreased potential for productivity in cryptophytes than in haptophytes or diatoms due to a smaller photosynthetic cross section (Claustre et al., 1997). A study examining cultured Arctic representatives of these three groups suggest a dependency on temperature; haptophytes only grew faster under higher temperatures (Van de Poll, unpublished data).

Few field-based studies have assessed the relationship between phytoplankton community composition and PP in the Southern Ocean (Uitz et al., 2009; Takao et al., 2012) or the WAP (Claustre et al., 1997; Vernet et al., 2008; Huang et al., 2012). All these studies observed a significant effect of species composition on water column PP. The most extensive study in the coastal WAP observed a positive relation between diatom abundance and water column PP (Vernet et al., 2008). In contrast, a negative relation was observed between cryptophytes and water column PP. Vernet et al. (2008) further estimated that phytoplankton species composition explained 11% of the variability in primary production. Yet relative phytoplankton species abundances are currently not used when modeling PP at the WAP. Estimates of daily depth-integrated PP over 1995–2006 along the WAP ranged from 250 to 1,100 mg C m⁻² d⁻¹. The coastal regions and Marguerite Bay were most productive, reaching maxima of ~5,000–7,200 mg C m⁻² d⁻¹ in highly productive years (Ducklow et al., 2012b; Weston et al., 2013; Stukel et al., 2015). A study modeling weekly PP measurements used a simple, but often used, model which fits their data reasonably and leaving room for improvement (Behrenfeld and Falkowski, 1997; Vernet et al., 2012). This model did not take the composition of the phytoplankton or macro nutrient availability into account, parameters previously known to be affected by the observed changes in climate. Investigating these relations will help tuning current global biogeochemistry models, which already incorporated a crude mechanism to describe phytoplankton community composition (e.g., Aumont et al., 2015).

This study aims to understand and model PP in a climatically variable, coastal region of the WAP where large changes in phytoplankton species composition and meltwater dynamics occur. Our main objective is to reassess the relations between PP, phytoplankton biomass, and species composition, light, macronutrient concentrations, and water column properties. Thereby, we study if parameters not traditionally used to model PP, such as variability in phytoplankton species composition and macro nutrient concentrations, would need to be included given rapid changes currently unfolding in the WAP and improve current models (Behrenfeld and Falkowski, 1997; Vernet et al., 2012). Most previously used models are appropriate for open ocean applications and not necessarily coastal systems. To this end, we used a non-assumptive model (Random Forest,

Breiman, 2001) based on *in situ* measurements to discern the contributions of different biological and environmental parameters governing PP. In addition, we seek a minimalistic but optimized model of PP unbiased by our previous understanding of phytoplankton productivity. This optimized model was then used to estimate PP during one austral summer in a region where PP was studied previously to ensure a good validation of our findings with independent studies. Finally, we seek to compare a statistical approach with a more traditional mechanistic method of estimating PP (Platt et al., 1980; Weston et al., 2013).

The basis for our modeling approaches was formed by field measurements collected at the British Antarctic Survey's Rothera Research Station in northern Marguerite Bay. Here, the year-round Rothera Oceanographic and Biological Time Series (RaTS) program, representing the larger northern Marguerite Bay region, provides the infrastructure and scientific context to understand the dynamics in phytoplankton productivity (Venables and Meredith, 2014). Previous studies have shown a variable, yet generally high, level of phytoplankton biomass dominated by diatoms during summers in this region (Annett et al., 2010; Kozłowski et al., 2011; Rozema et al., 2017a). Phytoplankton biomass, both total and per major taxonomic group, was shown to be related to summer water column stability which, in turn, was driven by winter sea ice cover (Venables et al., 2013; Rozema et al., 2017a). Typically, persistence of summer biomass appears to be regulated by wind mixing events, meltwater input, and nutrient availability (Piquet et al., 2011; Rozema et al., 2017b). The variability at this coastal site, where PP has occasionally been studied, will allow our models to cover a large variability in environmental conditions and phytoplankton

community dynamics. Also, the RaTS allows for sufficient context of our estimates on which we can build and validate our non-traditional modeling approach.

MATERIALS AND METHODS

Site, Sample Collection, and Rats Program

The sampling site was situated in northern Marguerite Bay near Rothera Research Station and is part of the RaTS-program (Figure 1; 67.570°S 68.225°W). Year-round observations of biological and physical processes at this site (15 m depth) started in 1997 (Clarke et al., 2008; Venables et al., 2013; Rozema et al., 2017a). The RaTS site is situated in water ~520 m deep and ~2 km from the nearest shore. Our extensive summer campaign (Nov 2013–Mar 2014) was run in parallel to the RaTS program. CTD and fluorescence measurements to a maximum depth of ~95 m were obtained using a live-feed from the CTD package (SeaBird 19+) equipped with an in-line fluorometer (WS3S, WETLabs), turbidity sensor measuring at 700 nm (ECO NTU, WETLabs), and a spherical sensor (SPQA, LICOR) for Photosynthetically Active Radiation (PAR). Multiple fixed (2, 15, and 75 m) and one variable (depth of fluorescence maximum; F_{max}) depths were sampled for pigment analysis by HPLC, macronutrients (N, P, and Si) and maximum quantum yield of photosystem II (PSII). Phytoplankton production through 14 C incorporation was measured at F_{max} . Surface oxygen isotopes (0 m) used in this study were part of the RaTS program supplemented with a small number ($n = 7$) of samples collected at 2 m. Desired sampling frequency during summer was twice per week, and once per week for photosynthetic parameters with

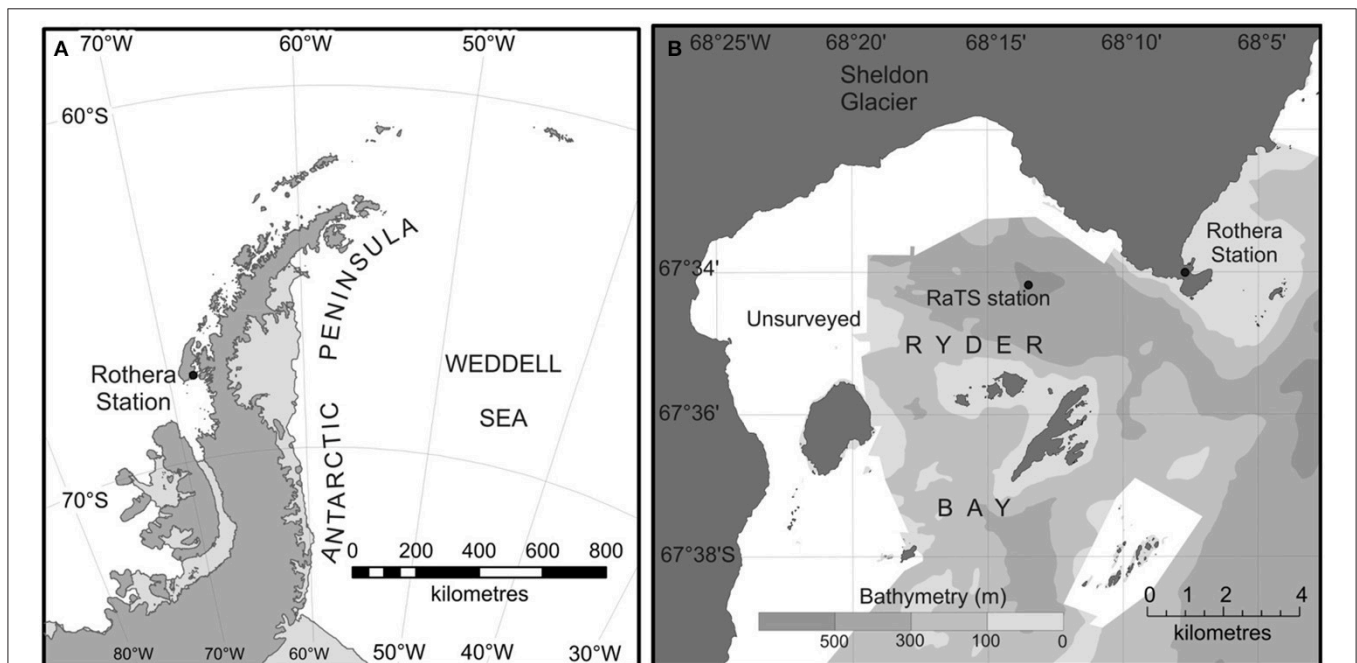


FIGURE 1 | (A): map of the western Antarctic Peninsula, marked with the location of Rothera Research Station on Adelaide Island in northern Marguerite Bay. **(B):** An expanded view of the immediate vicinity of Rothera, including Ryder Bay where the RaTS long-term monitoring station is situated.

^{14}C incubations (P-E). The fluorescence profiles were calibrated against Chl-*a* measured by HPLC and collected at 15 m and F_{max} . All samples were collected by deploying a 12–24 L Niskin bottle with a hand or electric winch on a small boat. After collection, the samples were transported in dark and insulated boxes to the laboratories for further processing. Daily wind speed data from Rothera station was obtained from the British Antarctic Survey (<http://www.antarctica.ac.uk/met/metlog/>).

PAR and Light Attenuation

Year-round measurements of PAR were collected every half hour at Rothera station using a fixed sensor (SKP215, Quantum Skye, UK). These measurements were integrated to estimate the daily irradiance dose at the surface ($\text{PAR}_{\text{daily}}$, in $\mu\text{mol photons m}^{-2} \text{s}^{-1}$). Attenuation in the water column (K_d in m^{-1}) was derived from linear regression of natural log-transformed, PAR-values as measured with the sensor on the CTD (Kirk, 1983). Estimates of K_d were based on the observations between 5 and 50 m unless PAR was below detection levels at depths shallower than 50 m. The first 5 m were excluded due to frequent shading by the sampling boat. Also, strong deviations in PAR within the water column during a cast, most likely due to changing cloud cover, resulted in a skewed estimate of K_d . Therefore, these casts were split and K_d was calculated by using the curve which had the majority of measurements. These estimates were linearly interpolated to obtain estimates for K_d on days between CTD casts and subsequently used to obtain daily light dose estimates for every depth for every half hour using (Equation 1):

$$E = \text{PAR}_{\text{daily}} \cdot e^{-K_d z} \quad (1)$$

where E ($\mu\text{mol quanta m}^{-2} \text{s}^{-1}$) is the daily irradiance dose at depth z (m) (Kirk, 1983).

Water Column Stability and Stable Oxygen Isotopes

Mixed layer depth (MLD) was defined using the depth at which the Brunt–Väisälä frequency (BVF) was highest, as recommended recently for the WAP (Carvalho et al., 2017). Ocean Data View v4.7.8 was used to calculate the BVF (Schlitzer, 2016). Oxygen isotope samples ($\delta^{18}\text{O}$) were analyzed as described in Meredith et al. (2008). We used the $\delta^{18}\text{O}$ and salinity data to derive quantitative estimates of the freshwater contributions from sea ice melt and meteoric water, which is possible since freshwater of meteoric origin is isotopically lighter than that originating from sea ice melt. The two sources of meteoric water are precipitation and glacial meltwater (Craig and Gordon, 1965). Simple budget calculations have shown that glacial discharge is the largest contributor to the meteoric water prevalence in the coastal WAP environment, however this glacial melt may be injected over a broader depth range than direct precipitation, which necessarily impacts the ocean directly at the surface (Meredith et al., 2017).

Macronutrients

Nutrient samples were collected as described earlier (Bown et al., 2017) or as described below. A cross comparison of these two different collection methods during the 2012–2013

season did not yield major differences. Subsamples (2×5 mL) for macronutrient analysis were filtered (prefilter: $0.8 \mu\text{m}$, membrane: $0.2 \mu\text{m}$, Acrodisc Supor PE, Pall, US) and stored in the dark at -20°C (nitrate, nitrite, and phosphate) or 4°C (silicate). Analyses for samples collected using both collection methods were conducted using a Technicon TRAACS 800 Auto-analyzer at the Royal Netherlands Institute for Sea Research, The Netherlands (Bown et al., 2017). Nitrite and nitrate were summed and are presented as N_{Tot} .

Phytoplankton Pigments

Collection for and measurement of phytoplankton pigments were done using the standard RaTS protocol (Rozema et al., 2017a). In short, phytoplankton cells were collected on GF/F filters (Whatman, US) under low light and at $\sim 2^\circ\text{C}$ using a mild vacuum (<0.2 mbar) generated with a water jet pump. After filtration, the filters were snap frozen in liquid nitrogen and stored at -80°C until analysis. For analysis, samples were freeze dried for 48 h prior to extraction in 90% acetone (v/v) at 4°C in the dark for another 48 h. The pigments were separated using a Waters 2,695 HPLC system equipped with a Zorbax Eclipse XDB-C8 column ($3.5 \mu\text{m}$ particle size) as described by van Heukelem and Thomas (2001) and modified by Perl (2009). Pigments were manually identified using retention times and diode array spectroscopy (type 996, Waters, US) before quantification. Calibration of the system was performed using standards (DHI LAB PRODUCTS, Denmark) for chlorophyll c_3 , peridinin, 19'-butanoyloxyfucoxanthin, fucoxanthin, neoxanthin, prasinoxanthin, 19'-hexanoyloxyfucoxanthin, alloxanthin, lutein, chlorophyll b, and Chl-*a*.

CHEMTAX (v1.95) was used to estimate the abundance of various phytoplankton groups (Mackey et al., 1996). This program employs a factor analysis and steepest descent algorithm to find the best fit using initial pigment ratios from previous studies for eight different phytoplankton classes, namely prasinophytes, chlorophytes, dinoflagellates, cryptophytes, two types of haptophytes, and two types of diatoms (Supplementary Table 1, Wright et al., 2009, 2010; Rozema et al., 2017a). Haptophytes were included as two separate groups since *Phaeocystis*, the most abundant haptophyte in our system, has shown great plasticity in cellular pigment ratios with iron and light availability (as reviewed in van Leeuwe et al., 2014). Also, two diatom groups were included to allow for differentiation in chlorophyll c_3 containing diatoms such as *Proboscia* spp. and *Pseudonitzschia* spp. and non- c_3 containing species (unpublished data).

A total of 139 samples were divided over 3 bins. We calculated from city-block distances and clustered the samples according to Ward's method as suggested by Latasa et al. (2010) using Past 2.17c (Hammer et al., 2001). This dendrogram suggested binning of the samples collected at 0–7, 8–49, and 50–75 m bins and the smallest bin size was 40. The initial ratios were as used previously with the RaTS data (Supplementary Table 1; Bown et al., 2017; Rozema et al., 2017a). Neoxanthin and prasinoxanthin were excluded for the 50–75 m bin. Further CHEMTAX settings were as described previously (Kozłowski et al., 2011; Rozema et al., 2017a). CHEMTAX was run 60 times

using randomized pigment: Chl-*a* ratios ($\pm 35\%$ of the initial matrix) to obtain the best possible result per bin (Supplementary Table 1; Wright et al., 2009). Both haptophyte and diatom groups as used with CHEMTAX were pooled and renamed to “pooled haptophytes” and “pooled diatoms,” respectively.

Estimating Phytoplankton Biomass

A quenching correction was applied to the upper water column fluorescence, as collected during the CTD casts, to allow for an accurate estimate of the water column biomass. This correction was based on linear regression of Chl-*a* concentration at 2 and 15 m or F_{\max} (the sample collected at the shallowest depth was used) as estimated from HPLC analysis. If F_{\max} was $>>15$ m (start summer campaign: late November 2013–Early December 2013) then Chl-*a* measured at F_{\max} and 2 m were used. The slopes of these regressions were then used to estimate biomass in relation to the calibrated CTD fluorescence for the top layer. We present the corrected phytoplankton biomass estimates as Chl-*a*_{corr}. Additionally, standing phytoplankton stocks were calculated by integrating the first 80 m of the water column using Chl-*a*_{corr}. The depth of 80 m was chosen as this is generally in the layer of winter water, near the deepest sample for HPLC (75 m) and light does not penetrate this deep during summer (Clarke et al., 2008).

Maximum Quantum Yield of PSII

Triplicate measurements of maximal quantum yield of PSII (F_v/F_m) using a WATER-FT PAM (Walz, Germany) in dark-adapted subsamples (40 mL) allowed for the assessment of the photophysiological state of the phytoplankton community (reviewed in Maxwell and Johnson, 2000). F_v/F_m -values < 0.5 are generally indicative of exposure to stress due to e.g., exposure to excess PAR and/or ultraviolet radiation or nutrient limitation.

Dissolved Inorganic Carbon

Dissolved Inorganic Carbon (DIC) concentrations were used in the calculation of primary production from ^{14}C measurements. Samples for the analysis of DIC were collected on 8 sampling days (January 15, 25, 30, February 3, 10, 17, 25, and March 3, 2014) in borosilicate glass bottles and analyzed within 20 h of sampling as presented in Jones et al. (2017). DIC concentrations were determined by coulometric analysis according Johnson and Sieburth (1987) and Dickson et al. (2007) using a VINDTA 3C instrument (Versatile INstrument for the Determination of Total Alkalinity, Marianda, Germany). A power function was used to describe the relationship between DIC and HPLC derived chlorophyll *a* concentrations ($R^2 = 0.94$, $p < 0.001$) to estimate DIC concentrations for the calculation of primary production on missing sampling days.

Photosynthesis-Irradiance Measurements

Water collected at the depth of F_{\max} was used to estimate phytoplankton productivity. PP was estimated by constructing P-E curves ($n = 16$) based on a modified protocol from Lewis and Smith (1983). Seawater subsamples (20 mL) were incubated in a photosynthetron for 4 h under 21 different light

intensities ($8\text{--}713 \mu\text{mol quanta m}^{-2} \text{s}^{-1}$; Osram Powerball HCL-TT 250W in a Jolly symmetrical armature) at *in situ* ($\pm 0.5^\circ\text{C}$) temperature after the addition of $0.37 \text{ MBq } ^{14}\text{C-bicarbonate}$ (PerkinElmer, The Netherlands; $>1.7 \text{ TBq mmol}^{-1}$) in clear 60 mL polyethylene terephthalate (PET) bottles (Qorpak, US). After incubation, the samples were filtered onto polycarbonate membrane filters (Millipore, US; 25 mm, $0.4 \mu\text{m}$ pore size) using a gentle vacuum (<0.1 bar) on a filtration carousel (Millipore, US). After filtration, the filters were acidified with fuming HCl for ~ 20 min to vent off unincorporated $^{14}\text{C-bicarbonate}$. Filters were placed into 20 mL PET scintillation vials (Wheaton, US) and supplemented with 10 mL scintillation cocktail (Ultima Gold, PerkinElmer, The Netherlands). For time zero activity, triplicate samples were prepared, filtered over polycarbonate membrane filters and immediately acidified. For total activity, 10 μL Ethanolamine was added to 100 μL ^{14}C spiked subsamples in triplicate in 20 mL PET scintillation vials containing a clean polycarbonate membrane filter. Time zero activity and total activity were then treated identically to the other samples. All samples were vortexed and left for 2 days to ensure maximum dispersal of activity throughout the scintillation cocktail before counting activity on a Tri-Carb 2900TR (Packard Instrument Company, US).

The data were converted to DPM after correction for time zero activity (Steeman-Nielsen, 1952), and normalized to Chl-*a*, as derived from the HPLC analyses, and DIC (see above), and corrected for metabolic discrimination (5%) between ^{12}C and ^{14}C (Ærtebjerg-Nielsen and Bresta, 1984). These data were fitted by least-squares nonlinear regression using the model Equation 2 of Platt et al. (1980), modified by MacIntyre and Cullen (2005):

$$P = P_s \left(1 - e^{-\alpha \left(\frac{E}{P_s} \right)} \right) \left(e^{-\beta \left(\frac{E}{P_s} \right)} \right) + P_0 \quad (2)$$

where P ($\mu\text{g C } \mu\text{g Chl-}a^{-1} \text{ h}^{-1}$) is the carbon fixation rate at irradiance E , P_s ($\mu\text{g C } \mu\text{g Chl-}a^{-1} \text{ h}^{-1}$) the light-saturated maximum of carbon fixation in the absence of photoinhibition, α [$\mu\text{g C } \mu\text{g Chl-}a^{-1} \text{ h}^{-1} (\mu\text{mol quanta m}^{-2} \text{s}^{-1})^{-1}$] the initial slope of the P-E curve, β [$\mu\text{g C } \mu\text{g Chl-}a^{-1} \text{ h}^{-1} (\mu\text{mol quanta m}^{-2} \text{s}^{-1})^{-1}$] is a measure for photoinhibition, and P_0 ($\mu\text{g C } \mu\text{g Chl-}a^{-1} \text{ h}^{-1}$) is the carbon uptake or release without any irradiance. These parameters were considered characteristic for the phytoplankton throughout the water column at the day of collection.

Constructing and Evaluate Models

Two different methods of calculating phytoplankton carbon uptake were employed. The classical, mechanistic approach relies on the estimations of the P-E parameters, biomass estimates and total irradiance for days without P-E incubations (M_{platt} ; Platt et al., 1980). These estimates were obtained as discussed below and run through M_{platt} for every 1 m. Our second statistical approach method uses Random Forest (RF) regressions. RF is a machine learning method which constructs a large number of regression trees by randomly taking subsets of the data and input variables (Breiman, 2001). The major benefits of using such a tool over more traditional (mostly linear) models are: Regression trees

allow for (1) non-linear relationships between input and output variables, (2) non-additive interactions between predictor and response variables and (3) correlations between different input variables (Breiman, 2001; Shi and Horvath, 2006). The relevant variables included were phytoplankton biomass, macronutrients, MLD, temperature, salinity, total irradiance and the relative abundance of the three most important phytoplankton groups, namely diatoms, haptophytes and cryptophytes (Behrenfeld and Falkowski, 1997; Arrigo et al., 1999; Dierssen and Smith, 2000; Vernet et al., 2008, 2012; Montes-Hugo et al., 2009; Piquet et al., 2011; Venables et al., 2013; Williams et al., 2016; Bown et al., 2017; Rozema et al., 2017a,b).

For the RF approach, we modeled the log of the PP estimates from the 16 P-E experiments (see above) using the depths for which all the aforementioned variables were available ($n = 62$). We constructed a model in which all the parameters (M_{RFmax}) were included and one reduced, optimized model (M_{RFmin}). During the construction we grew 2,000 trees, and four (M_{RFmax}) or two (M_{RFmin}) variables were used at every split. Variables least important in M_{RFmax} were removed individually until the fit of the new model no longer improved and this became M_{RFmin} . Partial dependence plots were generated for each variable per model to understand to the relationship with PP. These plots have the input parameter on the horizontal axis and give the range of impact of the input variable on PP. We discussed the importance of each variable to the models above but here we used the shape (and direction) of the curve to understand how this parameter influenced PP in our RF models. Partial dependence plots show the effect of the predictor on the response variable after accounting for the average effects of the other predictors. Therefore, it is the trend, not the actual values, that describes the dependence between both variables.

To assess the predictive power of both RF models, the data were split randomly into a training (80%) and test set (20%). This was repeated 10 times resulting in slightly different models of each type (M_{RFmax} and M_{RFmin}). These 10 slightly different models were subsequently tested against the randomly generated test sets using linear regression. We included the mean p and R^2 -values of these assessments. The “randomForest” package in R (v3.3.1) was used to build our models (Liaw and Wiener, 2002; R Core Team, 2016).

Constructing Input Matrices and Estimating Phytoplankton Productivity

After the construction of the minimal RF model, we aimed to calculate the PP in the water column (0–80 m) over the full course of the season using M_{platt} and M_{RFmin} . For this, we needed to obtain full matrices for all parameters used in the models. We used anisotropic ordinary kriging to interpolate both in depth and time using the “gstat” package in R (Pebesma, 2004). First, we optimized the kriging function to best fit the autocorrelation per variable for the depth component using variograms. This function was then used for the interpolation through time. We chose a weighing factor of 0.3 for the time component to allow for the required sensibility needed during

short, sudden events such as wind mixing. The matrix for PAR data was constructed as described above. Values for α , β , P_0 , and P_s were linearly interpolated from days with P-E incubations. Thus, identical biomass and PAR estimates were used for all modeling approaches.

PP was calculated for every half hour, due to the resolution of the light measurements, and every depth (0–80 m) using M_{platt} and M_{RFmin} . The values were summed to obtain daily values. We chose to use a resolution of half an hour as light was highly variable throughout the day and our model included the potential for photoinhibition. Secondly, our RF model included the aforementioned parameters and was run using the constructed matrices. For light in M_{RFmin} used the summed daily irradiance for all depths. We summed all data for December, January and February as this is traditionally the most productive period and define this as the summer productivity estimates.

RESULTS

Light

PAR availability in the water column was higher during spring (November) than during summer (Figure 2A). Also, PAR in the water column was lowest from December 15th to December 21th. This was related to high turbidity, increasing K_d to $>0.30\text{ m}^{-1}$ as well as low surface irradiance dose (December 23rd and 24th; Figures 2B,C). In contrast, surface irradiance dose peaked at the end of December before gradually decreasing. A brief and sharp increase of PAR at 15 m occurred on January 15th when K_d was low. Not until February 14th had PAR penetrated $>50\text{ }\mu\text{mol quanta m}^{-2}\text{ s}^{-1}$ beyond 15 m again and thereafter remained above this level (Figure 2A). The dynamics in K_d strongly coincided with standing phytoplankton biomass. The only major exception was after the short break (December 23rd and 24th) in our sampling effort due to hindrance of drifting sea ice sheets. During these casts, K_d was far higher ($>0.30\text{ m}^{-1}$) than observed at other times in our campaign.

Water Column Physics

Our field campaign started (November 26th) during a period where SSTs started to increase slightly while MLD and surface salinity decreased (Spring, Figures 3A,B, 4A). Above zero surface temperatures first occurred on and persisted onwards from December 6th (start of period 1). The MLD during the first half of December was shallow (1 m) while salinity kept decreasing. After a brief period of elevated wind speed and a deepening of the MLD (December 23rd), the water column surface salinity restabilized while the SST increased further (Figures 3A,B, 4A). Maximum SST at 1 m during this summer was 2.7°C on January 6th and 11th. The period of high SSTs was interrupted by an increase of wind speed and deepening of the MLD in mid-January ending period 1 and homogenizing water column temperature and salinity (Figure 4A). This increase in MLD was of a short duration as the water column restabilized within a week. Thereafter, surface temperature increased again with positive temperatures up to a depth of $\sim 30\text{ m}$ and a stable surface salinity (~ 32.6). These conditions lasted until mid-February when strong winds occurred more frequently, salinity started to

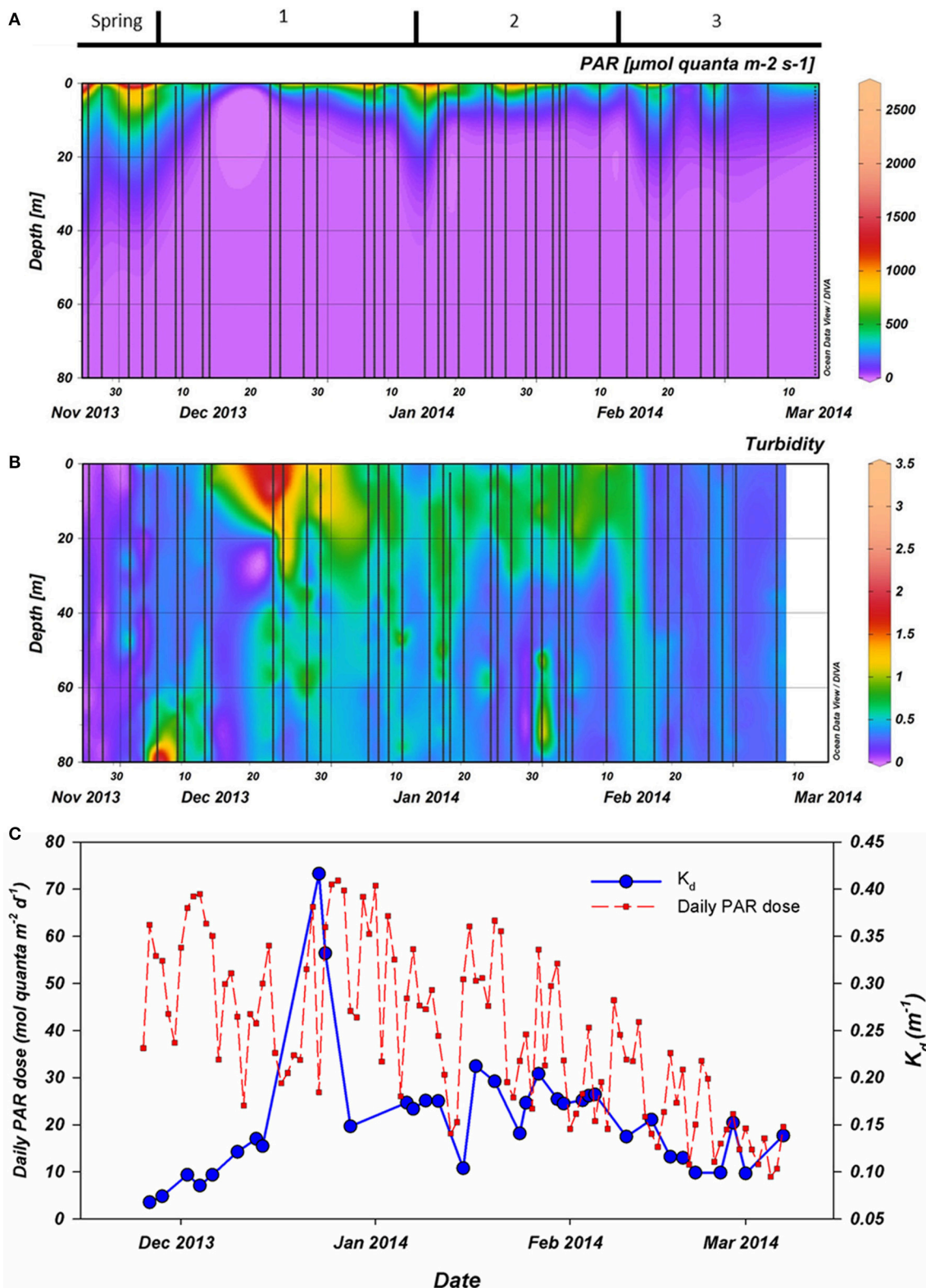


FIGURE 2 | (A): Photosynthetically active radiation (PAR) as measured during CTD casts (vertical casts). **(B):** Turbidity as also measured during the CTD casts. **(C):** First, the extinction coefficient (K_d in blue) as calculated using the PAR data collected by CTD. Also, the daily irradiance dose for PAR (red) integrated from continuous measurement at Rothera station. All data were collected during the 2013–2014 season. Period 1 starts on December 6th, period 2 on January 15th and period 3 on February 10th.

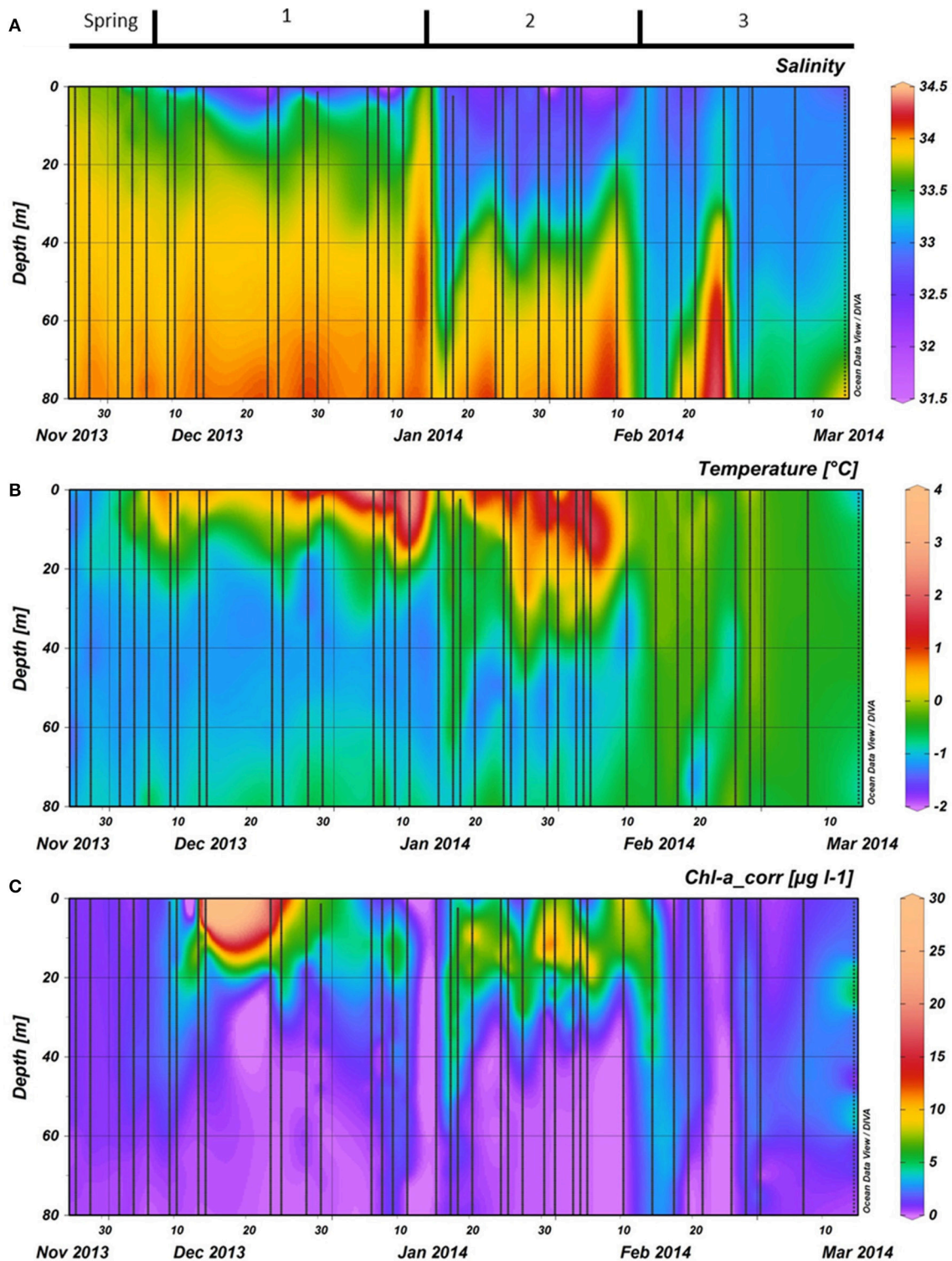
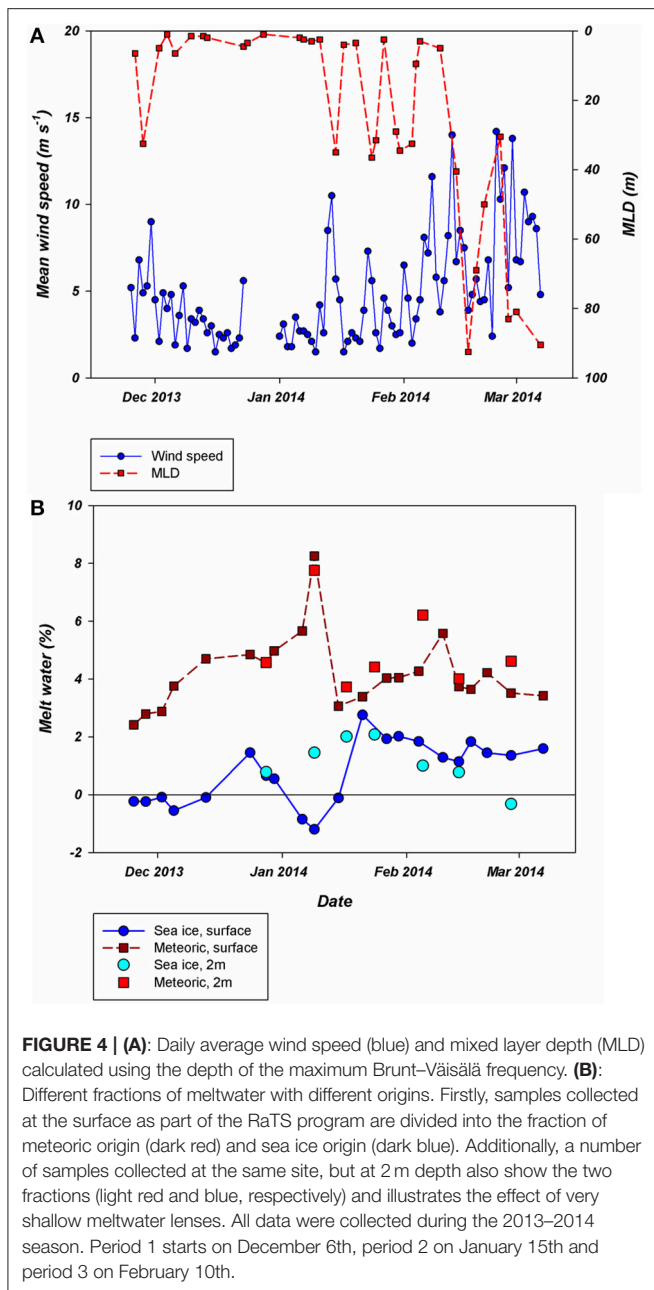


FIGURE 3 | (A): Salinity, **(B):** Temperature, and **(C):** Phytoplankton biomass as measured by fluorescence during the vertical CTD casts. Phytoplankton biomass fluorescence was calibrated against chlorophyll a measurements with HPLC and corrected for quenching in top layer using the chlorophyll estimates. All data were collected during the 2013–2014 season at the RaTS site. Period 1 starts on December 6th, period 2 on January 15th and period 3 on February 10th.



increase and SST decreased which resulted in the end of period 2. As in spring, the water column temperature dropped and stayed below 0°C and was more homogenous throughout the water column after February 10th, labeled as period 3.

Freshwater Origin

Sea ice retreat from the preceding winter was late and the contribution of sea ice meltwater to the water column during summer was, although modest at all times, the highest (2.8%) recorded in the RaTS program (Figure 4B; Legge et al., 2017; Meredith et al., 2017). Oxygen isotope data from the surface and at 2 m depth showed an influx of sea ice melt after December

13th, continuing to at least the end of February. The contribution of glacial melt was higher than sea ice melt (mean 4.4%), but also more variable. After an initial increase, freshwater of meteoric origin peaked at 8.3% (also a record for the RaTS program) before dropping to just 3.1% on January 15th as an increase in wind speed mixed the water column (Figure 4A). Both freshwater components increased rapidly after the period of MLD deepening, after which the meteoric freshwater contribution at 2 m depth remained relatively stable at ~4% while the contribution of sea ice melt slowly decreased toward the end of summer.

Macronutrients

Macronutrient concentrations at the end of spring were high with water column averages of 29.0, 1.8, and 83.5 μM for N_{Tot} , phosphate, and silicate, respectively (Figure 5). These values were characteristic of the Antarctic winter water, as concentrations at 75 m were only slightly higher. During period 1, the vertical distribution of the macronutrients strongly followed the patterns observed in SSTs. Nutrient concentrations were generally lowest at the surface (2 m). Nutrient concentrations in the first ~15 m strongly decreased over a 10-day period after December 13th. On December 23, all macronutrients were virtually depleted with only 0.01 μM phosphate, 0.06 μM N_{Tot} , and 0.70 μM silicate left at 2 m depth. This period of depletion lasted until the end of period 1, when nutrients throughout the water column were mixed, restoring 2 m values to the levels similar to those observed before December 13th. Low nutrient concentrations were observed in period 2 with minimum values at 2 m depth on February 5th (13.6 μM silicate, 0.07 μM nitrate and no detectable nitrate or phosphate). This depletion of nutrients persisted to a depth of ~20 m. Period 3 was characterized by a gradual increase in surface nutrient levels.

Phytoplankton Biomass and Species Composition

Phytoplankton biomass accumulated during period 1 (Figure 3C). Relative abundances of diatoms increased at the cost of haptophytes during the start of this period (Figure 6). A sudden increase in phytoplankton biomass (up to 26 μg Chl- a l^{-1}), mainly consisting of small diatoms (Buma et al. unpublished microscopy data; Figure 6, Supplementary Figure 1), coincided with the occurrence of sheets of sea ice in the bay on December 14th. This biomass peak lasted until December 28th after which values dropped to ~5 μg Chl- a l^{-1} . Integrated phytoplankton stocks (0–80 m) followed a similar trend: a large increase between December 10th and 14th, and a steady decrease thereafter (Figure 7). Moreover, F_v/F_m in the top 10 m during the short, yet sharp increase in biomass was lower than expected for the *in situ* light intensities when compared to the rest of the summer ($p < 0.001$; Supplementary Figure 2). After the biomass peak, phytoplankton stocks remained stable until the end of period 1. During period 2, diatoms remained the most important taxonomic group although cryptophyte proportions increased (max. 40%) in the surface layer (2 m; Figures 4A, 6). Also, haptophytes increased at 15 m (~15%) until January 30th. Phytoplankton biomass was more evenly distributed over the

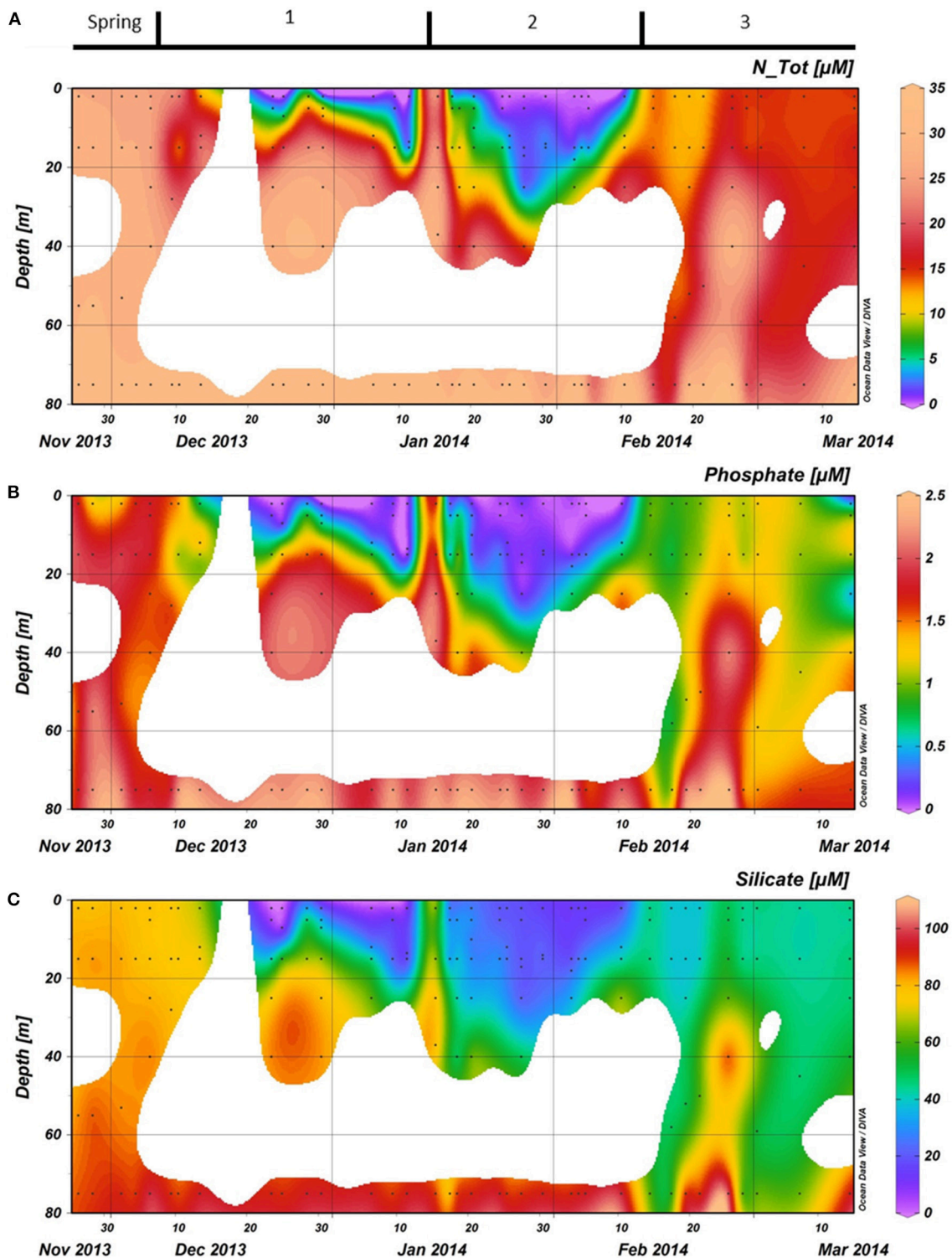


FIGURE 5 | (A): Total nitrogen (nitrate + nitrite), **(B):** Phosphate, and **(C):** Silicate concentrations in the water column at the RaTS site. Black dots illustrate the sampling depths. Period 1 starts on December 6th, period 2 on January 15th and period 3 on February 10th.

first 30 m during this period, reaching a maximum concentration of $11 \mu\text{g Chl-a l}^{-1}$ and suggested three short periods where biomass increased and decreased (Figure 7). The first was

associated with the increase in haptophytes at 15 m, the second and third with diatoms. In period 3, relative abundances of haptophytes and cryptophytes were both elevated in comparison

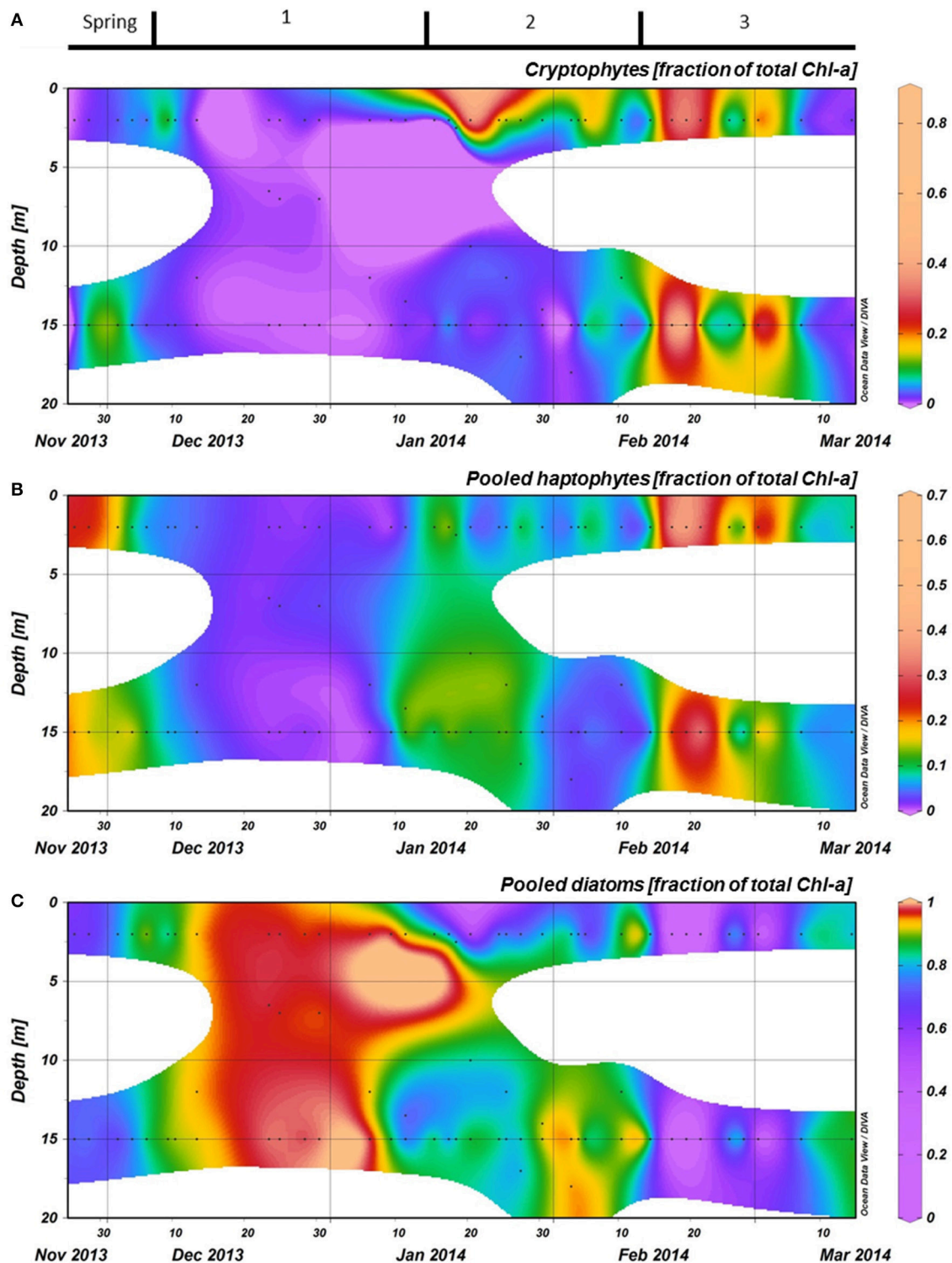


FIGURE 6 | The relative abundances of the three most important phytoplankton groups, as calculated with CHEMTAX using HPLC measurements of pigments, are shown for the top 20 meters at the RaTS site. **(A):** Pooled diatoms, **(B):** Pooled haptophytes, and **(C):** Cryptophytes. Black dots illustrate the sampling depths.

to the previous periods (max. 38 and 47%, respectively) and both phytoplankton groups co-occurred in the first 15 m (Table 2). Diatoms were more abundant at the deep fluorescence maxima.

Phytoplankton biomass at 75 m was low and only increased during the two occasions where strong winds caused a deepening of the MLD. Relative group abundance matched those observed

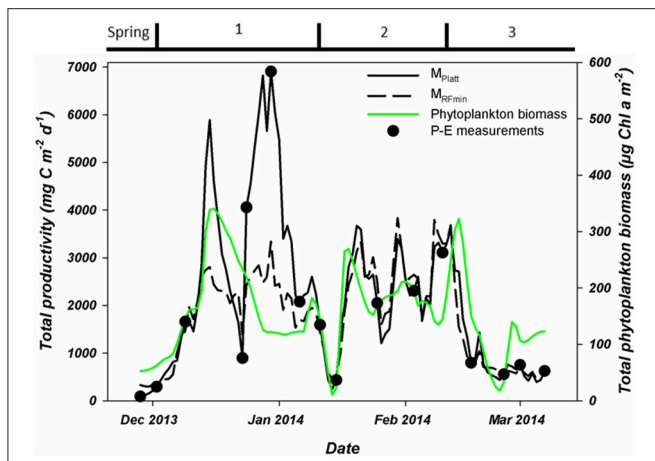


FIGURE 7 | Total, water column integrated primary production as measured during P-E experiments (black dots) or estimated using two modeling approaches. These estimates are based on Platt et al. (1980; solid black line) or the minimal random forest model constructed in this study (dashed black line). Also shown is the water column integrated phytoplankton biomass. All integrations were to a depth of 80 meters. Period 1 starts on December 6th, period 2 on January 15th and period 3 on February 10th.

in the surface, albeit with a delay of ~ 30 days (Supplementary Figure 1).

P-E Measurements

Variability in photosynthetic parameters as obtained from the P-E experiments ($n = 16$) changed over the course of the summer. At the end of spring (November–early December), α , P_s , and P_0 were low in comparison to the summer values (Table 1). At the end of December, both α and P_s increased $> 50\%$. P_s appeared to reach an average of $\sim 4 \mu\text{g C } \mu\text{g Chl-a}^{-1} \text{ h}^{-1}$ for the rest of the summer (January–March) while P_0 increased further to $0.68\text{--}0.85 \text{ C } \mu\text{g Chl-a}^{-1} \text{ h}^{-1}$. The α also increased further to $0.068\text{--}0.083 \mu\text{g C } \mu\text{g Chl-a}^{-1} \text{ h}^{-1} [\mu\text{mol quanta m}^{-2} \text{ s}^{-1}]^{-1}$. In contrast, β was very low and only increased to a maximum average of $0.004 \mu\text{g C } \mu\text{g Chl-a}^{-1} \text{ h}^{-1} [\mu\text{mol quanta m}^{-2} \text{ s}^{-1}]^{-1}$, suggesting that photoinhibition was of minor importance.

Using these P-E parameters, we calculated integrated PP for the water column on the days of the experiments (Figure 7). These values ranged from 92.5 (end of November) to $6,907.7$ (end of December) $\text{mg C m}^{-2} \text{ d}^{-1}$. While the PP estimates from these experiments showed an increasing trend over the entire month of December, PP on December 23rd was substantially lower. The peak value of $6,907.7 \text{ mg C m}^{-2} \text{ d}^{-1}$ occurred on December 30th after which PP decreased reaching a minimum of $438.2 \text{ mg C m}^{-2} \text{ d}^{-1}$ on January 15th. PP during period 2 increased again to a maximum of $3,109.5 \text{ mg C m}^{-2} \text{ d}^{-1}$ and lasted until mid-February before stabilizing around $680 \text{ mg C m}^{-2} \text{ d}^{-1}$ during period 3.

Testing Importance of Species Composition

To estimate the potential influence of phytoplankton species composition on PP we compared the log PP at discrete depths

where we collected supplementary data (if $\text{PP} > 0$, $n = 42$) against the relative abundances of the three major phytoplankton groups (Table 2). It appeared that while PP and both diatoms and haptophytes approached a monotonic relation, cryptophytes and PP did not. Linear regressions for these three comparisons yielded r^2 -values of 0.01 , 0.27 , and 0.14 ($p < 0.001$) for cryptophytes, haptophytes and diatoms, respectively. Thus, we opted for spearman rank order analyses, which are less sensitive for non-linear trends (Table 2). We observed a negative relation between PP and haptophytes whereas a larger diatom fraction resulted in higher PP ($p < 0.001$). However, the relation between cryptophytes and PP was no longer significant ($p = 0.36$). The opposing trends between haptophytes and diatoms vs. PP were supported by correlations between the phytoplankton groups (Table 2). Haptophytes and cryptophytes were positively related to each other while both were negatively related to diatom abundance ($p < 0.001$).

Creating RF Models

Using M_{RFmax} with all the parameters included ($p < 0.001$, $R^2 = 0.91$), we quantified the contribution of the different input parameters on PP using a variable importance plot (Figure 8). Of these parameters, daily PAR was the most important, increasing the mean square error (MSE) by 50.4% . The other two large components were phytoplankton biomass (26.7%) and water temperature (21.6%). The three macronutrients (N_{Tot} , phosphate and silicate) showed a smaller importance ($20.4\text{--}17.3\%$). Salinity (15.5%), MLD (13.6%) and the relative contributions of the major phytoplankton species ($7.3\text{--}12.2\%$) influenced the MSE to a lesser extent. The contribution of some of the 12 included parameters was limited, possibly because some of the variability was already explained by a correlated and more important parameter. Therefore, we reduced our first model (M_{RFmax}) to obtain a more minimalistic model (M_{RFmin}) with only three parameters that explain 93% of the variability: daily irradiance dose throughout the water column, phytoplankton biomass and N_{Tot} (Figure 8). The second-best option explained only slightly less of the MSE and included seawater temperature instead of N_{Tot} . The inclusion of either seawater temperature or N_{Tot} resulted in highly similar minimal models. A linear correlation analysis on all the samples in which N_{Tot} was measured (excluding 75 m samples) revealed a strong relation between the two parameters ($p < 0.001$, $r^2 = 0.67$).

Partial Response of Variables

After the construction of our models, we analyzed how each of these parameters influenced PP at the depths where all auxiliary data (generally 2 , 15 , 75 m, and F_{max}) were available using partial dependence plots (Figure 9). The partial dependence plots for the two different models were highly similar and thus we present M_{RFmax} as it includes all variables, those for M_{RFmin} are included in the supporting information (Supplementary Figure 3). Light was the most important factor in both models, the partial dependence plot suggests that when light is not available, PP is strongly decreased. Also, when light is above a certain threshold ($\sim 3 \text{ mol quanta m}^{-2} \text{ d}^{-1}$), its effect on PP remains unchanged. Phytoplankton biomass showed a similar trend. PP

TABLE 1 | Shown below are the mean values (\pm standard deviation) for photosynthetic parameters P_s and P_0 (both in $\mu\text{g C } \mu\text{g Chl-a}^{-1} \text{ h}^{-1}$), and α and β [both in $\mu\text{g C } \mu\text{g Chl-a}^{-1} \text{ h}^{-1} (\mu\text{mol quanta m}^{-2} \text{ s}^{-1})^{-1}$] as calculated from the ^{14}C experiments at F_{max} .

	<i>n</i>	Chl-a	DIC _{calc}	P_s	P_0	α	β
Nov–Dec*	3	1.1 \pm 0.9	1.1 \pm 0.9	1.347 \pm 1.177	0.486 \pm 0.438	0.040 \pm 0.027	0.000 \pm 0.000
Dec [#]	3	12.6 \pm 4.8	12.6 \pm 4.8	3.803 \pm 3.409	0.569 \pm 0.501	0.059 \pm 0.047	0.002 \pm 0.003
Jan	4	4.6 \pm 2.9	4.6 \pm 2.9	3.521 \pm 0.589	0.797 \pm 0.435	0.080 \pm 0.030	0.002 \pm 0.002
Feb	4	4.1 \pm 2.4	4.1 \pm 2.4	4.228 \pm 1.174	0.633 \pm 0.592	0.068 \pm 0.040	0.004 \pm 0.004
Mar	2	2.80 \pm 0.3	2.80 \pm 0.3	3.950 \pm 0.191	0.847 \pm 0.454	0.082 \pm 0.026	0.001 \pm 0.002

Also presented are Chl-a ($\mu\text{g l}^{-1}$) as a proxy for phytoplankton biomass and DIC_{calc} ($\mu\text{mol kg}^{-1}$) during these experiments. The values below are averages of *n* experiments conducted between Nov 2013 and Mar 2014. Only 1 experiment was run at the end of November and was pooled with the early December values (up to the 9th and indicated by *). The values for December 24 or later are therefore shown separately (as indicated by #). We did not use these averages for our calculations but the measurements per time point.

TABLE 2 | Spearman rank order correlations where log PP estimates (*n* = 42) for the depths for which pigment samples were collected (with *PP* > 0) were correlated to the calculated relative abundance of the major phytoplankton groups as calculated with CHEMTAX.

	Log PP	Rel. cryptophytes	Rel. haptophytes	Rel. diatoms
Log PP	–			
Rel. cryptophytes	–0.15	–		
Rel. haptophytes	–0.65*	0.57*	–	
Rel. diatoms	0.57*	–0.70*	–0.93*	–

The used PP estimates were from the days on which a photosynthesis vs. irradiance experiment was conducted. Significant correlations are indicated by *.

did not increase further above $\sim 7 \mu\text{g Chl-a l}^{-1}$, although the support of this plateau is limited given that it is based on only 10–20% of the data. The three macronutrients had similar trends in relation to PP: the lower the concentration, the higher the PP until it reaches a plateau. Silicate and N_{Tot} dynamics suggest two distinct plateaux in its relation to PP, coinciding with the concentrations in the two different bloom periods (before and after January 15th). Seawater temperature showed a strong bimodal effect where 0°C was pivotal, likely related to the freeze of thaw cycle of glacial and sea ice. In contrast, a decreasing salinity and MLD coincided with a gradual increase in PP. Finally, the contribution of relative species abundances show a strong positive effect on the presence of diatoms as the effect on PP is strongly reduced below $\sim 93\%$ diatoms. Roughly 70% of our data have decreased diatoms abundances suggesting a strong support for this observation. As such, increasing fractions of cryptophytes and/or haptophytes are associated with decreased PP.

Estimating Summer PP

PP over the full course of the summer season was estimated using two different approaches, the traditional M_{Platt} and our best RF model, M_{RFmin} (Figures 7, 10). Dynamics of both PP estimates were very similar (Figure 10), and both methods produced near identical values for the period after January 15th (periods 2 and 3). While the patterns in PP for December and early January (period 1) were similar, the magnitude of PP differed greatly (Figure 10). At most, PP estimated in December was a factor 2.5 higher using M_{Platt} than when calculated using M_{RFmin} .

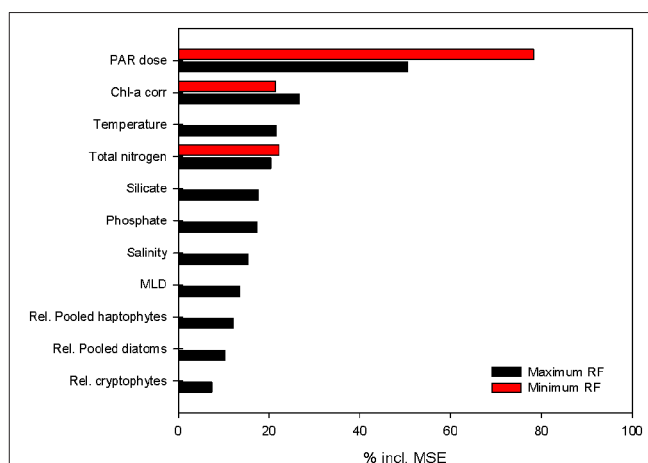


FIGURE 8 | Variable importance plot for the two random forest models build to estimate PP throughout the water column: R_{Fmax} (black) and R_{Fmin} (red). The x-axis shows the percentage of reductions in mean square error (MSE) per variable. All variables were included in the maximum model; daily PAR dose in the water column, phytoplankton biomass (chlorophyll a corrected for quenching), temperature, total nitrogen, silicate, phosphate, salinity, mixed layer depth (MLD based on maximum Brunt–Väisälä frequency) and relative abundance of the three most abundant phytoplankton groups. The minimum model excluded variables until a model was reached with the highest possible fit. The importance of the three remaining variables in this model is shown in red.

On average, M_{Platt} was 40% higher during the first month of the summer reaching a maximum of $6,908 \text{ mg C m}^{-2} \text{ d}^{-1}$ on December 31st (Figure 7). PP in December peaked twice in both models, namely on December 15th and 31st. After January 15th, three steep increases in PP occurred on January 21st, 30th, and February 8–12th. The maxima of these periods were of the same magnitude at those calculated for December with the M_{RFmin} model. In contrast, using M_{Platt} PP in January and February was only half of the values observed in December. A second noteworthy difference between the two methods of estimation is to which depth PP occurs (Figure 10). However, these differences contribute little to the summed PP (Figure 10). Total summer PP (Dec–Feb) was estimated to be 214.4 g C m^{-2} using M_{Platt} and 176.1 g C m^{-2} using M_{RFmin} , or 2.68 and $1.96 \text{ g C m}^{-2} \text{ d}^{-1}$, respectively.

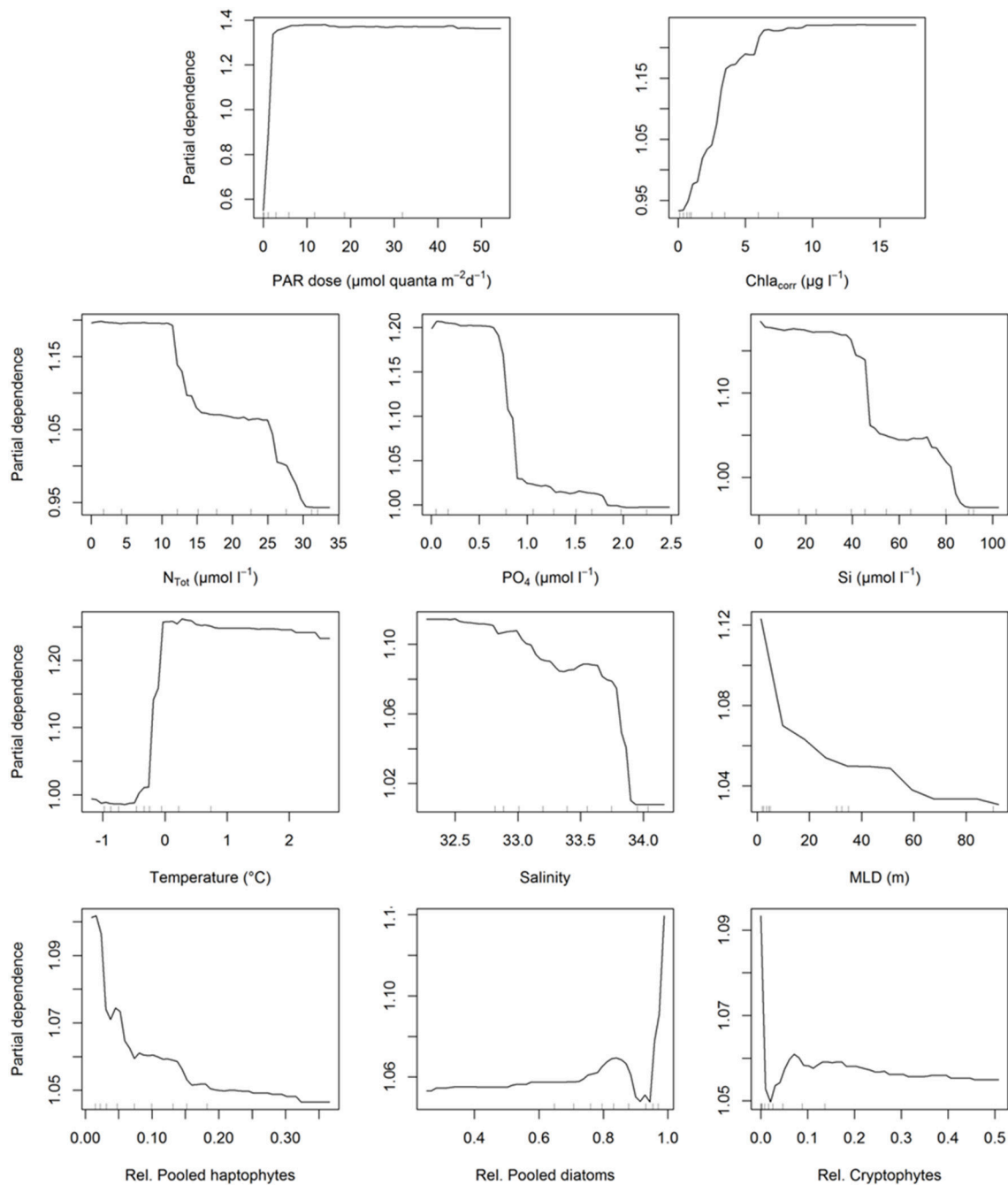


FIGURE 9 | Partial dependence plots for all the variables included in the maximum random forest model to assess the importance of each value in the modeling of PP. The partial dependence plots for the minimal model are shown in Supplementary Figure 3. Inner tick marks represent the 10% cohorts.

DISCUSSION

In the present study we measured PP at a coastal monitoring site over one full summer and modeled PP using two approaches (mechanistic and statistical), resulting in three different models (M_{Platt} , M_{RFmax} , and M_{RFmin}). Firstly, we briefly describe how our field site and the summer season dynamics were

representative of the wider WAP region and conditions, both past, present and future. Then, we evaluate the M_{RFmax} model to find relations between environmental parameters and PP. The four most important parameters in relation to PP were PAR, phytoplankton biomass, seawater temperature and N_{Tot} . This full model was simplified through the reduction in parameters to obtain the minimal representative model capable

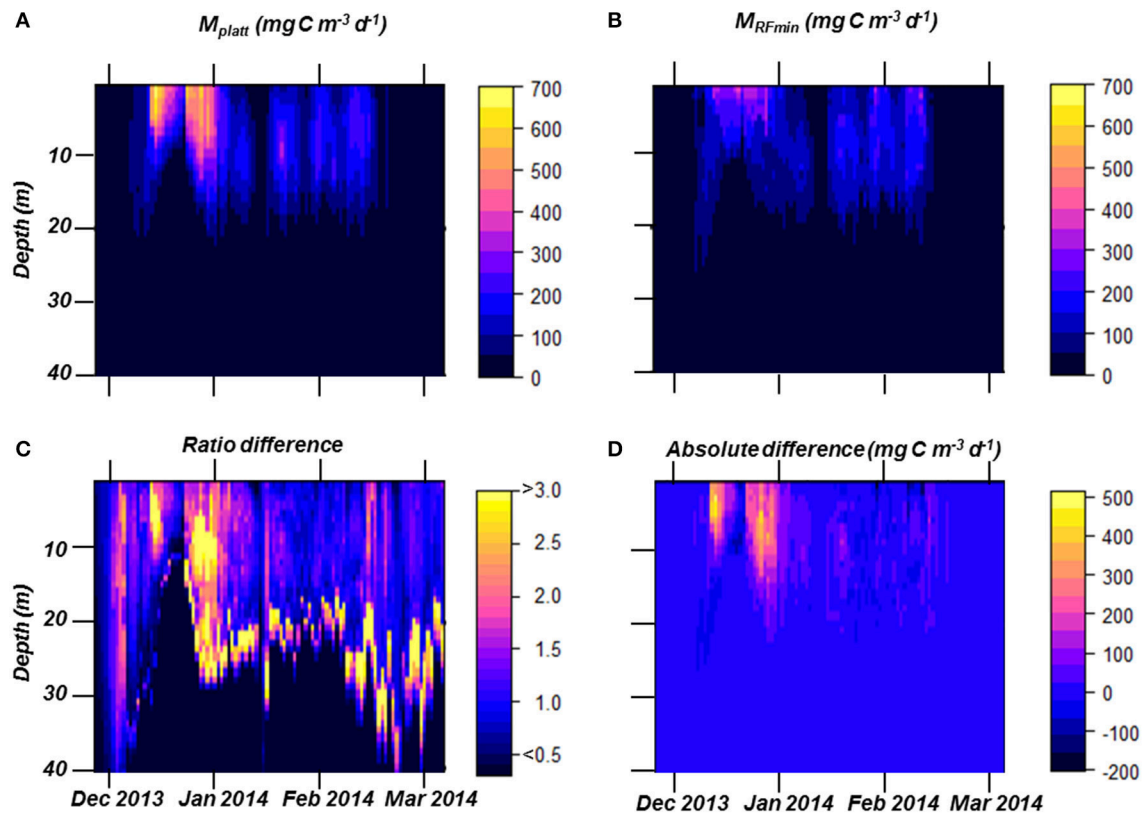


FIGURE 10 | Estimated primary production (PP) in the top 40 m based on M_{Platt} (A) and on M_{RFmin} (B). The ratio of PP between the two models (C), values >1 indicate a higher estimates using M_{Platt} . This ratio is used to understand the differences in behavior between the two models. Finally, we show M_{Platt} minus M_{RFmin} (D) to present when and at which depths the two models differ or match.

of predicting PP, M_{RFmin} . The minimal model only included PAR, phytoplankton biomass and N_{Tot} . Finally, we compare both models and discuss the implications of our M_{RFmin} on the current efforts of modeling phytoplankton production in the coastal WAP.

From our *in situ* measurement, we identified four different periods during the 2013–2014 summer season (Figures 2, 4; spring, 1, 2, and 3), which were divided by the influx of large sheets of sea ice (period 1) or wind mixing events (ending periods 1 and 2). This suggests an adequate covering of past, current and future conditions in the coast WAP (Ducklow et al., 2007, 2013). Period 1 was strongly influenced by sea ice whereas 2 and 3 were not. Accurately covering of these distinctively different periods is important given the continuously decreasing expand of winter sea ice (Stammerjohn and Maksym, 2017). This summer could be classified as one of medium productivity (median summer biomass at 15 m was $4.3 \mu\text{g Chl-a l}^{-1}$), slightly elevated SSTs and typical sea ice dynamics (Meredith et al., 2012; Venables et al., 2013; Rozema et al., 2017a). Below we briefly review the characteristics of these different periods and how these are representative for other coastal sites along the WAP to establish the range of conditions to which our models were exposed.

Influence of Presence/Absence of Sea Ice on Phytoplankton Dynamics

During period 1, the sea ice gradually melted and freshened the surface layer thereby reducing MLD and promoting diatom growth, as is typical for the traditional WAP ecosystem (Ducklow et al., 2012a, 2013). Moreover, a strong increase of turbidity indicated the marked presence of non-photosynthetic material, presumably microalgal aggregates released from the bottom of the sea ice (Figure 2B; Legge et al., 2017). Low F_v/F_m -values and relatively low values for α further suggests that the diatoms in these aggregates were low light acclimated which is typical for the sea ice environment (Table 1, Supplementary Figure 2, Arrigo et al., 2014). The difference in F_v/F_m between the late December community and the rest of the season suggests that the late December community was of a different origin, presumably the sea ice, confirming that period 1 could be deemed representative for the traditional coastal phytoplankton dynamics.

Classical features of the sea ice environment were the sole dominance of diatoms and depleted silicate stocks, the latter unexpected given the coastal nature of the site and never documented prior to the 2013–2014 summer (Clarke et al., 2008; Annett et al., 2017; Bown et al., 2017; Cassarino et al., 2017; Rozema et al., 2017b). Even though macronutrient

concentrations in the water column during this period were low and uptake by phytoplankton was likely hampered, nutrients could still have been available through high bacterial remineralization, sloppy grazing, and/or viral lysis which peaks after the end of the bloom (Ducklow et al., 2012b; Brum et al., 2015). Additionally, the build-up of phytoplankton biomass could also have been ended by strong grazing, as is often observed at the sea ice edge, an attribute not covered in the RaTS program (Ross et al., 2008; Saba et al., 2014; Steinberg et al., 2015).

Brief wind mixing events marked the starts of both periods 2 and 3, two periods uninfluenced by sea ice cover. These mixing events reconfigured the water column, lower SST, and mixed the freshwater to a greater depth, thus changing the conditions for phytoplankton growth, as evident from changes in the community composition and PP. In period 2, the water column was restabilized by a fresh and shallow surface layer, which was populated by cryptophytes which are often associated with glacial meltwater (Moline et al., 2004; Saba et al., 2014). Below this layer, haptophytes and later large diatoms (mostly *Proboscia* spp.) thrived. Period 3 was characterized by increased wind mixing causing a more homogenous water column and deeper MLDs, typical for conditions at the end of summer and observed more frequently at the WAP (Clarke et al., 2008; Montes-Hugo et al., 2009; Venables and Meredith, 2014; Rozema et al., 2017b). Thereafter, the community consisted of large proportions of haptophytes, diatoms and cryptophytes (Figure 6).

Evaluation of Parameters Potentially Impacting PP

First and foremost, our statistical model showed the importance of light availability and biomass in the promotion of PP. While completely expected, it does establish further confidence in our statistical model, which needed to establish these relations from the training set as opposed to the mechanistic model where these relations were already established.

Nutrient concentrations during summer can be highly variable and even undetectable near the surface (Figure 5; Clarke et al., 2008; Bown et al., 2017; Rozema et al., 2017b). Governing of PP (or biomass accumulation) by macronutrients in coastal Antarctic regions have been described previously and suggested a potential limitation of nitrogen (Alderkamp et al., 2012b; Henley et al., 2017; Rozema et al., 2017b). Moreover, the nutrient dynamics are strongly influenced by the source of meltwater. Glacial melt is low in macronutrients and organic matter and dilutes the surface layer (Henley et al., 2017). This opposed to sea ice melt, a prominent source of organic matter and bacteria to remineralized the nutrients (as reviewed in Fripiat et al., 2017). Additionally, deep (wind) mixing replenishes the nutrient concentrations of the euphotic zone. These three different components control MLD in the coastal WAP system and explains why nitrogen is a better predictor for PP than MLD (Meredith et al., 2008; Rozema et al., 2017b). The positive effects of a low salinity, above 0°C water temperature and shallow MLD on PP support the importance of glacial melt in shaping the PP. But in this study, these parameters did not contribute significantly in predicting PP (Figure 8).

Moreover, our partial dependence plots suggest negative relationships between nutrient concentrations and PP (Figure 9) when above 12, 0.7, and 42 μM for N_{Tot} , phosphate and silicate respectively and neutral when below these thresholds. We consider these negative relations an effect from the phytoplankton community becoming dominated by only one (or at least very few) species for which the conditions are optimal when the community is blooming (Annett et al., 2010). The blooming species, which contributes most to PP, takes up most of the macro nutrients and dominates. We suspect that the blooming species draws down nutrients to concentrations in line with their uptake kinetics and half-saturation constants and, when reaching these minimum concentrations, form the plateaus in the relations between macronutrients and PP (Huisman and Weissing, 1999; Timmermans et al., 2004).

When we consider all the parameters included in our M_{RFmax} model, the added benefit of including phytoplankton community composition is limited (Figure 8). However, this does not imply that Chl-a specific production does not vary between the groups in the natural assemblages. The random forest model selects the parameters that explain the statistically highest possible amount of the MSE of PP. Yet, some of the variability explained by the top parameters is also be explained by mechanisms related to lower ranking parameters. Thus, while adding those parameters would not improve the model this does not mean that they are not ecologically or mechanistically important. For example, literature suggests that haptophytes favor a more unstable water column and our results confirm such a relation (Arrigo et al., 1999; Rozema et al., 2017a). However, haptophyte relative abundance is suggested to be of only minor importance in relation to PP by our models, despite a large difference between photosynthesis strategies between haptophytes and e.g., diatoms (Kropuenske et al., 2009; Alderkamp et al., 2012a). However, as we know haptophytes are related to deeper MLDs, and therefore lower SSTs, higher macronutrient concentrations and more variable PAR (Kozłowski et al., 2011; Rozema et al., 2017a). Thus, most of the variability in PP explained by knowing the fraction of haptophytes is already captured by the aforementioned parameters that rank higher. As such, we conclude that adding species information would not significantly improve models of PP in the coastal system. Sufficient knowledge of the physical, (photo)chemical properties and total phytoplankton biomass in the water column will suffice for predicting PP.

PP Estimations

Calculation of PP by both approaches (M_{Platt} and M_{RFmin}) was strongly similar except during the second half of December (Figure 10, period 1). The most important difference between these ~ 3 weeks and the rest of the season was the presence of melting sea ice. Estimates using M_{Platt} were higher (53%) during period 1 than those using M_{RFmin} . However, integrated biomass during period 1 was not exceptionally high when compared to period 2 or 3. In fact, it was lower than expected given the PP estimates from both models. Apparently, the high PP as measured using the P-E parameters did not result in an increase in total phytoplankton biomass. A difference in Chl-a specific production between taxonomic groups is

unlikely to have been played a major role given the low contribution of taxonomic information to PP in M_{RFmax} (Figure 9). The large discrepancy between productivity and phytoplankton biomass accumulation could be due to a number of reasons. (1) Phytoplankton released from the sea ice are low light acclimated and were lowering their Chl-a:carbon ratio while acclimating to higher irradiance intensities in the water column, thus growth would remain undetected using Chl-a as we used Chl-a as a proxy (Sakshaug and Holm-Hansen, 1986). (2) Phytoplankton Chl-a:carbon ratios remain relatively unchanged, but the phytoplankton mainly replenish their micro- and macronutrient reserves before biomass accumulation by growth (Arrigo, 2005). These two explanations are supported by the lower F_v/F_m , possibly indicative of photo inhibition, and low α observed for this population (Table 1, Supplementary Figure 2). Additional explanations are related to ecosystems dynamics. They assume that the high productivity is translated into phytoplankton biomass, but there is also a large loss factor. (3) Sea ice occurrence is often associated with krill and other grazing which can efficiently lower phytoplankton biomass (Behrenfeld, 2010). (4) Viral lysis as after a period of biomass increase, viruses often lyse which include the bursting of phytoplankton cells and thus lowering biomass (Brum et al., 2015). Finally, the last and possibly most likely option (5) would be fast sedimentation of microalgal aggregates dominated by heavily silicified diatoms (Riebesell et al., 1991). The last three theories are supported by the observation of a very high turbidity in the surface layer (Figure 2B), not occurring elsewhere in the summer despite similar phytoplankton biomass concentrations. Our fieldwork cannot conclusively exclude any of the five hypotheses mentioned above or combinations thereof, that created the observed dynamics in phytoplankton biomass and proportions thereof to PP during period 1. Most likely it is a combination of multiple of the aforementioned explanations. Thus, while M_{RFmin} resembles mechanistic models that estimate PP in the marine environment, it would greatly benefit from the inclusions of parameters better explaining the phytoplankton dynamics at the sea ice edge. Also, the relation between N_{Tot} and PP does provide extra information for future efforts modeling PP in the coastal Antarctic ocean.

Our two estimates of 176.1 and 214.4 g C m⁻² per summer are in fair agreement with multiple different estimates of PP from previous years. Two recent estimates of net PP of 146 g C m⁻² yr⁻¹ for 2009–2010 and an estimate of 192 g C m⁻² yr⁻¹ for 2005–2006 for the RaTS site are comparable to ours (Weston et al., 2013; Henley et al., 2017). The latter estimate also employed P-E experiments with a very similar set up to ours. A slightly lower productivity of 99.4 g C m⁻² over a 5 month period was observed at a more northerly site near Palmer Station, generally considered less productive than northern Marguerite Bay (Stukel et al., 2015; Rozema et al., 2017a). Furthermore, our estimates are in broad agreement with a multi-year (1995–2006) analysis of PP using ¹⁴C along the WAP (Vernet et al., 2008). Another study based on the 2007–2008 summer but also covering the large WAP region estimate a lower productivity (78 g C m⁻² over 4 months) although this estimate is from a low biomass summer (Huang et al., 2012; Rozema et al., 2017a). Another estimate

derived from calculating NCP from silica isotopes conducted at the RaTS, has calculated a maximum of only 56.4 g C m⁻² over 4 months in 2009–2010 (Annett et al., 2017). Such a large discrepancy might be related to the bias of a silica based method to diatoms as haptophytes and cryptophytes also contribute to the carbon drawdown (Figure 9) and diatom abundance was low in the 2009–2010 seasons (Rozema et al., 2017a).

Implications and Perspectives

Our data underline the importance of sea ice for phytoplankton biomass accumulation and PP in the coastal seas. Thus, a decrease in sea ice, as has been observed for the WAP since the start of the satellite record, will have substantial impact on the coastal marine system (period 1; Stammerjohn et al., 2008). Also, the diatoms associated with the sea ice and the related meltwater layer are pivotal in the food web due to the importance in the krill life cycle (Flores et al., 2012). While the loss of sea ice would result in a decrease in phytoplankton, the influence of meltwater from glaciers is rising (period 2; Moline et al., 2004; Montes-Hugo et al., 2009). While this meteoric meltwater also promotes stratification, which in turn promotes PP, the timing of the establishment of glacial meltwater layers and the phytoplankton communities within are different potentially affecting the entire food web (Moline et al., 2004; Montes-Hugo et al., 2009; Mendes et al., 2013; Saba et al., 2014; Rozema et al., 2017a). While most PP models are designed for ocean regions, the MLD at the coastal ocean is established differently than in the open ocean due to the proximity of glaciers thus are generally not adequately included in these models.

Our minimal model also underlines that dissolved nitrogen and seawater temperature can be used to estimate PP. While we do not infer a causal relation between PP and these parameters, we observe strong relation to PP. Our RF does not fully explain the dynamics in PP, namely only 93%, and we consider this is related to the dynamics occurring in December. The strong reduction in phytoplankton biomass despite a high PP suggests another source which influences phytoplankton stocks, as discussed above. We consider the counterintuitive relation between N_{Tot} and PP evidence of important underlying processes not captured by our accurate yet simple model. Thus, a future improvement would be the measurement and incorporation of these undescribed processes into a model. Also, the near binary effect of above and below 0°C sea surface temperatures only indicates the importance of meltwater and mixing to the winter water layer. No relation to PP was observed with increasing seawater temperature while the strength of the relation between PP and salinity declines strongly when approaching lower values (<32.5).

CONCLUSIONS

We presented a phytoplankton PP model in the highly productive northern Marguerite Bay region. We thereby considered the effect of variability in relative phytoplankton group abundances to be adequately covered by the implementation of physical and (photo)chemical properties of the water column. Firstly, we show the challenge of modeling PP in phytoplankton communities

from the sea ice. This underlines the importance of the effort to obtain measurements from (below) the sea ice. Secondly, nitrogen concentrations were a stronger predictor of PP than MLD. Possibly because of the large difference in nitrogen concentrations between glacial and sea ice melt, the main sources for salinity driven stratification. Finally, our statistical models show that relative species abundances do not have to be included in future modeling efforts while maintaining a high level of confidence in the model. This last conclusion greatly simplifies the monitoring efforts required to provide field based measurements needed to estimate PP. The combination of these insights, based on both a classical, mechanistic model and independent, statistical models, should help shape the priorities for modeling efforts to accurately estimate PP in the climatically-sensitive and variable coastal WAP region.

AUTHOR CONTRIBUTIONS

PR, GK, AB, and WvdP have designed the field experiments and protocols. MM leads the RaTS program which supplied data to this study, and helped with interpretation of the isotope data. PR has performed the field experiments and data collection. PR, MV, WvdP, and GK have designed and/or performed the data analyses. PR wrote the manuscript with input from AB and WvdP. All authors contributed to a discussion of the manuscript and approved the final outcome.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fmars.2017.00184/full#supplementary-material>

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